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## Weed Species Associated with Cacao Cultivation Exhibiting Phytoremediation Potential for Cadmium and Lead: A Study Based on Bioconcentration and Translocation Factors

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**ABSTRACT:** The exploration and identification of spontaneous weed species in heavy metal-contaminated soils represent a relevant approach for understanding the role and their potential application in phytoremediation. In cacao cultivation, cadmium contamination poses a significant risk due to the restrictions established for soils and cacao-derived products, thereby threatening productive sustainability and export viability. The objective of this study was to identify weed species associated with cacao cultivation exhibiting accumulation patterns and phytoremediation potential for Cd and Pb, through the assessment of biomass production and the bioconcentration factor (BCF) and translocation factor (TF), in natural conditions. Soil samples were collected from seven cacao-growing zones, and the site with the highest Cd concentration ( $1.65 \text{ mg kg}^{-1}$ ) was selected, where ten dominant weed species were identified and evaluated. Significant interspecific differences were observed in biomass production and in Cd and Pb concentrations ( $p < 0.05$ ). *Talinum paniculatum* exhibited the highest total biomass ( $6.15 \text{ g}$ ) and the highest TF for Cd, whereas *Cyperus aggregatus* showed the greatest total Cd accumulation ( $2.68 \text{ mg kg}^{-1}$ ). BCF and TF values enabled the identification of species with accumulation patterns consistent with Cd phytoextraction (*T. paniculatum*, *C. aggregatus*, *Pseudelephantopus spiralis*, *Bidens riparia*, *Euphorbia heterophylla*, and *Malvastrum coromandelianum*) and with phytostabilization (*Ocimum campechianum*). For Pb the observed patterns were consistent with potential phytoextractive behavior in *B. riparia*, *P. spiralis*, *E. heterophylla*, *Hillieria latifolia*, and *Acalypha stachyura*. Although these results do not constitute functional evidence of remediation, they describe heavy metal accumulation and translocation patterns in native weed species, providing a technical basis for the design of future experimental trials aimed at the sustainable remediation of contaminated soils.

**KEYWORDS:** Spontaneous weeds; cadmium; lead; plant biomass; bioconcentration factor; translocation factor; rhizosphere

## 1 Introduction

The contamination of agricultural soils with heavy metals, particularly cadmium (Cd) and lead (Pb), represents a significant risk to human health and food security [1]. Cadmium can be readily absorbed by plants and translocated from the soil to plant tissues depending on its bioavailability, which is influenced by a range of edaphic and environmental factors [2]. In contrast, Pb has no known biological function, and its entry into the food chain occurs primarily through irrigation practices, environmental contamination during field cultivation, and post-harvest handling and packaging of agricultural products [1]. Under natural conditions, soils generally contain low Cd concentrations; however, its bioavailability is strongly governed by soil pH, organic matter content, and texture [3], and can be substantially enhanced by agricultural practices such as phosphate fertilization and pesticide application [4,5].

This issue has been widely documented in cacao (*Theobroma cacao* L.) cultivation, where Cd availability in soils promotes its uptake and translocation to the beans [6,7], in some cases reaching concentrations higher than those detected in the soil itself [8]. In response to the increasing risks associated with heavy metal contamination and the growing demand for cacao-derived products, maximum permissible limits have been established for both soils and cacao beans. The Ministry of the Environment of Peru has established Environmental Quality Standards with maximum concentrations of  $1.4 \text{ mg kg}^{-1}$  for Cd and  $70 \text{ mg kg}^{-1}$  for Pb in agricultural soils [9]. Regarding cacao-derived products, regulatory limits include  $0.6 \text{ mg kg}^{-1}$  for ground cacao [10] and  $0.7 \text{ mg kg}^{-1}$  for chocolate with a cocoa solids content  $\geq 30\%$  and  $< 50\%$  [11].

According to projections by Thomas et al. [12], high concentrations of Cd in Peruvian cocoa beans and soils generally affect at least 20% of producing households from six regions, and 86% in Piura region. The regulations have raised concern among producers, as a substantial proportion of national cacao production is destined for international markets, with at least 75% [13–15]. Argüello et al. [14] reported that, to prevent cacao beans from exceeding Cd concentrations of  $1 \text{ mg kg}^{-1}$ , soil Cd levels should not surpass  $0.4 \text{ mg kg}^{-1}$  under acidic conditions (pH 5.0), whereas at neutral pH (7.0) the threshold may increase to as much as  $1.0 \text{ mg kg}^{-1}$ .

Against this backdrop, the development of technologies aimed at mitigating heavy metal contamination in agricultural soils is essential [16,17], in order to reduce carcinogenic risks and adverse effects on human health [18]. Within this context, phytoremediation has emerged as a low-cost and environmentally sustainable alternative [19], based on the ability of certain plant species to extract, immobilize, or transform soil contaminants [20], while also producing substantial biomass, adapting to adverse conditions, and exhibiting a high capacity for metal accumulation [21].

A wide range of plant species has been evaluated for the phytoremediation of contaminated soils, including woody species [22,23], agricultural crops [24,25], as well as grasses and other herbaceous plants [26]. However, the effectiveness of these species is often constrained by site-specific environmental and edaphoclimatic factors [27], highlighting the need to identify plant species adapted to each agroecosystem [28]. In this regard, weeds are characterized by aggressive growth, native origin, high adaptability, and tolerance to adverse conditions [29–31]. Their physiological and genetic traits confer remarkable ecological plasticity [32], positioning them as promising candidates for the phytoremediation of soils contaminated with Cd and Pb [33,34].

Despite these advances, a significant knowledge gap persists regarding the exploration and identification of weed species with phytoremediation potential associated with cacao cultivation, an agroecosystem that covers more than 250,000 ha in Peru. Within this context, the objective of this exploratory study was to identify and evaluate the phytoremediation potential of spontaneous weed species adapted to the edaphoclimatic conditions of cacao-growing systems in the central Peruvian rainforest. This prospective

approach provides a technical foundation for the development of future controlled trials aimed at more precisely defining the role of these species in mitigating heavy metal contamination in agricultural soils.

## 2 Materials and Methods

### 2.1 Study Area

The research was conducted in the district of San Martín de Pangoa, Satipo Province, Junín Region, Peru. The area corresponds to the tropical premontane pluvial forest (bp-PMT) life zone, according to the Holdridge World Life Zone Classification System [35]. The site is characterized by maximum and minimum temperatures of 31.6 and 19.3°C, respectively, a relative humidity of 81.5%, and a mean annual precipitation of 1898 mm year<sup>-1</sup> [36]. One of the main economic activities in the area is the cultivation of coffee and cacao for both domestic consumption and export markets.

### 2.2 Experimental Design

An exploratory observational study was conducted based on purposive field sampling. Seven cacao-growing areas were identified according to the following criteria: plots at the production stage, a minimum area of one-hectare, organic management, manual weed control, and the absence of weeding activities during the two months prior to sampling. To determine soil heavy metal concentrations and identify the area with the highest Cd levels, as well as to characterize its physicochemical properties, soil sampling was carried out following the approach proposed by Havlin et al. [37]. This area was selected to maximize the likelihood of identifying species with metal accumulation capacity, consistent with the exploratory nature of the study. Soil samples were collected at a depth of 30 cm from edaphically homogeneous areas, with three replicates obtained per study site.

Soil samples were analyzed within the Soil, Water, and Foliar Laboratory Network of the National Institute for Agrarian Innovation (LABSAF-INIA). Prior to physicochemical analyses and the determination of Cd and Pb concentrations, samples underwent a pre-treatment consisting of air-drying at temperatures below 40°C and sieving to obtain the <2 mm fraction, in accordance with the procedure established by the International Organization for Standardization [38].

The variables considered for soil characterization included the following parameters and reference methodologies: sand, silt, and clay contents determined by the Bouyoucos hydrometer method [39]. Soil pH measured in a 1:1 soil–water suspension [40]; electrical conductivity (EC) measured in a 1:5 soil–water extract [41]. Organic matter content determined by the Walkley and Black method [39]; total nitrogen measured by the micro-Kjeldahl method [42]. Available phosphorus in neutral and acidic soils determined using the Bray and Kurtz method [39]. Available potassium determined according to Bazán Tapia [43]; exchangeable bases (Ca<sup>2+</sup>, Mg<sup>2+</sup>, K<sup>+</sup>, and Na<sup>+</sup>) extracted with ammonium acetate. Moreover, exchangeable acidity (H<sup>+</sup>, Al<sup>3+</sup>) extracted with potassium chloride [39]. Finally, effective cation exchange capacity (ECEC) was calculated as the sum of exchangeable bases and exchangeable acidity.

Total recoverable Cd concentrations were determined following the USEPA Method 3050B [44]. The resulting solutions were analyzed using flame atomic absorption spectrometry (FAAS). Standard quality assurance and quality control (QA/QC) procedures were applied throughout the analytical process. A calibration coefficient (R<sup>2</sup>) of 0.9995 or higher was obtained for the standard calibration curve on the flame atomic absorption spectrophotometer prior to sample analysis. Following initial calibration, the calibration curve was verified using an initial calibration blank (ICB) and an initial calibration verification standard (ICV). The ICV was prepared from an independent certified reference material (second source) at or near the mid-range of the calibration curve, with acceptance criteria set at ±10% of the true value. Spike

recovery of Cd from a certified reference material (90%–110%) and duplicate analyses performed every 10 samples were also included as part of the QA/QC protocol. The method detection limit for Cd using FAAS was 0.41 mg kg<sup>-1</sup>.

Soil analysis results indicated that the area known as Ciudad de Dios exhibited the highest Cd concentration (1.65 mg kg<sup>-1</sup>), exceeding the Peruvian Environmental Quality Standard (EQS) for agricultural soils established by the Ministry of the Environment of Peru [9]. Accordingly, this site was selected for the identification and analysis of spontaneous weed species associated with cacao cultivation.

### 2.3 Characterization of the Selected Study Area

The selected area comprises an 8-year-old cocoa plantation with a planting density of 3 m × 3 m, arranged in a triangular (staggered) pattern, and previously used for coffee cultivation. The plantation was classified as polyclonal, predominantly composed of CCN-51 (80%), followed by ICS-1 (5%), ICS-6 (5%), TSH-565 (3%), VRAE-15 (2%), VRAE-99 (2%), ICS-39 (1%), ICS-95 (1%), and other minor genotypes (1%). Agronomic management included pruning carried out as required and fertilization based on seabird guano and dolomite (Fertiphos Tropical), resulting in an average productivity of 1000 kg ha<sup>-1</sup>. The presence of major diseases and pests was observed, including witches' broom (*Moniliophthora perniciosa*), frosty pod rot (*Moniliophthora roreri*), cocoa pod borer (*Carmenta foraseminis*), and brown spot disease (*Phytophthora* sp.).

Soil analysis indicated a clay loam texture, a high organic matter content (4.1%), and a moderately acidic pH of 6.10. Electrical conductivity was low (0.09 dS m<sup>-1</sup>), while total N, available P, and exchangeable K contents were 0.21%, 6.48 mg kg<sup>-1</sup> and 77.80 mg kg<sup>-1</sup>, respectively. The cation exchange capacity (CEC) was 3.50 cmol (+) kg<sup>-1</sup>, with exchangeable cation concentrations of Ca<sup>2+</sup> (2.84), Mg<sup>2+</sup> (0.56), K<sup>+</sup> (0.08), and Na<sup>+</sup> (0.02). In addition, to determine heavy metal (Cd and Pb) levels in cocoa plants, samples of leaves, pods, husks, and cotyledons were collected. Sampling followed a zigzag pattern, collecting plant material from ten trees, including five mature leaves with petioles and two mature pods per tree, free of visible disease or pest damage [45].

In the laboratory, leaves were washed with distilled water; cocoa pods were opened, cut into small fragments, and placed in beakers, while cocoa beans underwent a five-day fermentation process followed by shade drying. Leaves, pods, and cocoa beans were subsequently oven-dried at 70°C for 48 h until constant dry weight was achieved [46]. The husk was manually separated from the cotyledons, and each plant component was ground using porcelain mortars (Table 1).

**Table 1:** Cd and Pb concentrations in cocoa plant tissues.

| Sample         | Cd (mg kg <sup>-1</sup> ) | Pb (mg kg <sup>-1</sup> ) |
|----------------|---------------------------|---------------------------|
| Leaf           | 2.05                      | 4.75                      |
| Cocoa pod husk | 0.93                      | 2.08                      |
| Shell          | 0.92                      | 1.45                      |
| Cotyledon      | 0.89                      | 1.81                      |

### 2.4 Weed Sampling and Taxonomic Identification

For plant sampling, a systematic survey was conducted across the select plot in which dominant weed species were defined as those exhibiting the highest frequency of occurrence and visual cover within the evaluated plots. Based on this criterion, the selected species were collected, ensuring that sampled individuals per species were spatially distributed across different sectors of the field, thereby avoiding

clustering in a single location. Each plant individual constituted an independent experimental unit, allowing the acquisition of biological replicates per species, and was processed individually for subsequent analyses.

Species-specific percentage cover was estimated across the study area, after which dominant weed species were randomly collected with ten replicates at the flowering stage [28]. These ten replicates were obtained to ensure a representative sampling, provide sufficient plant material for laboratory analyses, and support accurate taxonomic identification. From these, three individual plants per species were selected and analyzed independently for heavy metal determination. Samples consisted of both aboveground and root tissues [47]. In addition, targeted rhizospheric soil samples were collected for heavy metal analysis [28]. The ten most representative species throughout the study area were collected and deposited in herbarium presses to facilitate dehydration and prevent the growth of microorganisms, such as fungi [48]. Taxonomic identification was performed according to the APG IV system The Angiosperm Phylogeny Group [49], at the Herbarium of the Department of Biology, Universidad Nacional Agraria La Molina, Lima, Peru.

## 2.5 Parameter Evaluation

### a. Weed biomass

Samples were washed with distilled water to remove adhering soil residues. Subsequently, the aboveground and root portions were separated. Each sample was oven-dried separately at 70°C until a constant weight was achieved [46]. The dried samples were ground using a stainless-steel mill and passed through a 100-mesh sieve [50]. Biomass was expressed on a dry weight basis, including aboveground biomass, root biomass, and total biomass, calculated from three replicates per weed species.

### b. Bioconcentration factor (BCF) and translocation factor (TF)

The bioconcentration factor (BCF) indicates the ability of plants to remove chemical components from the soil [32]. BCF is defined as the ratio between the total concentration of a metallic element in plant tissues and its concentration in the soil [51]. BCF values < 1 indicate metal-excluding species, whereas BCF values > 1 indicate metal-accumulating species [52,53]. This factor was calculated as follow:

$$BCF = \frac{\text{Concentration in the plant organ (mg kg}^{-1}\text{)}}{\text{Concentration in soil (mg kg}^{-1}\text{)}}$$

The translocation factor (TF) determines the ability of a plant to transfer a compound from the roots to other plant organs [51]. A plant is considered to exhibit phytostabilization potential when metals are not translocated to other organs and remain immobilized in the root system (TF < 1) [54]. In contrast, TF values > 1 indicate the plant's capacity to translocate metals from roots to aboveground tissues, such as shoots, and are therefore indicative of bioaccumulation potential [55]. This factor was calculated using the following equation:

$$TF = \frac{\text{Concentration in the aboveground part (mg kg}^{-1}\text{)}}{\text{Concentration in the root (mg kg}^{-1}\text{)}}$$

## 2.6 Statistical Analysis

Statistical analyses were performed using R software version 4.5 through the RStudio interface (2024 edition). Cd and Pb concentrations, as well as biomass, were modeled using generalized linear models (GLMs) implemented with the *glmmTMB* package. Model selection was conducted using the *MuMIn* package based on the corrected Akaike Information Criterion (AICc). Model fit was evaluated



using  $R^2$  as implemented in the *performance* package. Assumption checking included residual normality, outlier detection, and homogeneity of variances, which were assessed using the *DHARMA* package. Multiple comparisons of heavy metal concentrations and biomass among weed species were performed using the *emmeans* package, considering a significance level of  $\alpha = 0.05$  and applying false discovery rate (FDR) correction.

Additionally, the relationships between soil metal concentrations and biomass with metal contents in weed species were modeled using generalized linear models (GLMs) with spline functions (using different degrees of freedom and polynomial orders), implemented through the *splines* package. Predicted values were obtained using the *ggeffects* package, and results were visualized using the *ggplot2* package.

### 3 Results

#### 3.1 Identified Weed Species

The weed species recorded in the study area were distributed across eight botanical families. The identified species included *Pseudelephantopus spiralis* (Less.) Cronquist, *Ocimum campechianum* Mill., *Bidens riparia* Kunth, *Tripogandra serrulata* (Vahl) Handlos, *Euphorbia heterophylla* L., *Talinum paniculatum* (Jacq.) Gaertn., *Hillieria latifolia* (Lam.) H. Walter (HIW), *Cyperus aggregatus* (Willd.) Endl., *Acalypha stachyura* Pax, and *Malvastrum coromandelianum* (L.) Garcke (Table 2). These species are locally known as achicoria, wild basil, chilco, siempre viva, leche-leche, verdolaga, cuerda de indio, pata de gallina, cola de zorro, and pichana, respectively.

**Table 2:** Identified weed species.

| Species                                             | Family        |
|-----------------------------------------------------|---------------|
| <i>Pseudelephantopus spiralis</i> (Less.) Cronquist | Asteraceae    |
| <i>Ocimum campechianum</i> Mill.                    | Lamiaceae     |
| <i>Bidens riparia</i> Kunth                         | Asteraceae    |
| <i>Tripogandra serrulata</i> (Vahl) Handlos         | Commelinaceae |
| <i>Euphorbia heterophylla</i> L.                    | Euphorbiaceae |
| <i>Talinum paniculatum</i> (Jacq.) Gaertn.          | Talinaceae    |
| <i>Hillieria latifolia</i> (Lam.) H. Walter         | Petiveriaceae |
| <i>Cyperus aggregatus</i> (Willd.) Endl.            | Cyperaceae    |
| <i>Acalypha stachyura</i> Pax                       | Euphorbiaceae |
| <i>Malvastrum coromandelianum</i> (L.) Garcke       | Malvaceae     |

#### 3.2 Cadmium in Weed Species

The evaluated weed species showed significant differences in biomass production and Cd concentration ( $p < 0.05$ ) (Table 3). *T. paniculatum* recorded the highest total biomass (6.15 g) and root biomass (3.72 g), and ranked second in shoot biomass (2.43 g). The rhizospheric soil associated with this species exhibited a Cd concentration of  $0.91 \text{ mg kg}^{-1}$ , ranking fourth among the evaluated species. In plant tissues, *T. paniculatum* showed a total Cd concentration of  $0.78 \text{ mg kg}^{-1}$ , with  $0.59 \text{ mg kg}^{-1}$  in shoots, and was notable for presenting the lowest Cd concentration in roots ( $0.19 \text{ mg kg}^{-1}$ ). In contrast, *M. coromandelianum* exhibited the highest shoot biomass (3.30 g); however, it was associated with the third-highest Cd concentration in soil ( $1.05 \text{ mg kg}^{-1}$ ) and ranked fourth in shoot Cd concentration ( $1.04 \text{ mg kg}^{-1}$ ).

In contrast, *T. serrulata* exhibited the lowest total biomass (0.57 g) and root biomass (0.10 g), and ranked among the species with the lowest shoot biomass (0.48 g). Nevertheless, this species grew in soils with the third-highest Cd concentration ( $1.05 \text{ mg kg}^{-1}$ ) and showed some of the lowest Cd concentrations in plant tissues, including total Cd ( $0.66 \text{ mg kg}^{-1}$ ), root Cd ( $0.42 \text{ mg kg}^{-1}$ ), and shoot Cd ( $0.24 \text{ mg kg}^{-1}$ ).

Conversely, *C. aggregatus* was recorded in soils with the highest Cd concentration in soil ( $1.49 \text{ mg kg}^{-1}$ ) and exhibited the highest values of total Cd accumulation in plant ( $2.68 \text{ mg kg}^{-1}$ ), as well as the highest root ( $1.04 \text{ mg kg}^{-1}$ ) and shoot Cd concentrations ( $1.64 \text{ mg kg}^{-1}$ ). However, this species ranked third in total biomass (3.20 g), second in root biomass (1.68 g), and fourth in shoot biomass ( $1.52 \text{ g}$ ).

**Table 3:** Cadmium concentration in soil, roots, and aerial parts, and biomass by weed species: ANOVA, model selection, and adjusted means.

| Factors                    | Cd Soil<br>( $\text{mg kg}^{-1}$ ) | Cd Plant<br>( $\text{mg kg}^{-1}$ ) | Cd Root<br>( $\text{mg kg}^{-1}$ ) | Cd Shoot<br>( $\text{mg kg}^{-1}$ ) | Total<br>Biomass (g) | Root<br>Biomass (g) | Shoot<br>Biomass (g) |
|----------------------------|------------------------------------|-------------------------------------|------------------------------------|-------------------------------------|----------------------|---------------------|----------------------|
| Species                    | <0.01                              | <0.01                               | <0.01                              | <0.01                               | <0.01                | <0.01               | <0.01                |
| Model selection            |                                    |                                     |                                    |                                     |                      |                     |                      |
| Distribution               | Gamma                              | lognormal                           | lognormal                          | Lognormal                           | Normal               | lognormal           | lognormal            |
| AICc                       | -69.81                             | -46.77                              | -65.05                             | -61.36                              | -44.41               | -66.29              | -72.01               |
| R <sup>2</sup>             | 0.99                               | 0.99                                | 0.99                               | 0.99                                | 0.99                 | 0.99                | 0.99                 |
| Estimated means by species |                                    |                                     |                                    |                                     |                      |                     |                      |
| BRK                        | 1.22b                              | 1.79c                               | 0.59c                              | 1.20c                               | 0.69g                | 0.17e               | 0.52g                |
| CAE                        | 1.49a                              | 2.68a                               | 1.04a                              | 1.64a                               | 3.20c                | 1.68b               | 1.52d                |
| EHL                        | 1.50a                              | 1.87b                               | 0.9b                               | 0.97e                               | 0.99f                | 0.15e               | 0.85f                |
| MCG                        | 1.05c                              | 1.92b                               | 0.88b                              | 1.04d                               | 3.69b                | 0.39d               | 3.30a                |
| OCM                        | 0.30e                              | 0.65e                               | 0.42d                              | 0.22g                               | 2.18d                | 0.59c               | 1.58c                |
| PSC                        | 1.20b                              | 1.93b                               | 0.59c                              | 1.35b                               | 1.60e                | 0.58c               | 1.02e                |
| TPG                        | 0.91d                              | 0.78d                               | 0.19e                              | 0.59f                               | 6.15a                | 3.72a               | 2.43b                |
| TSH                        | 1.05c                              | 0.66e                               | 0.42d                              | 0.24g                               | 0.57h                | 0.10f               | 0.48g                |

Note: Different letters indicate significant differences among species ( $p < 0.05$ ). PSC: *Pseudelephantopus spiralis* (Less.) Cronquist; OCM: *Ocimum campechianum* Mill; BRK: *Bidens riparia* Kunth; TSH: *Tripogandra serrulata* (Vahl) Handlos; EHL: *Euphorbia heterophylla* L.; TPG: *Talinum paniculatum* (Jacq.) Gaertn; CAE: *Cyperus aggregatus* (Willd.) Endl; MCG: *Malvastrum coromandelianum* (L.) Garcke.

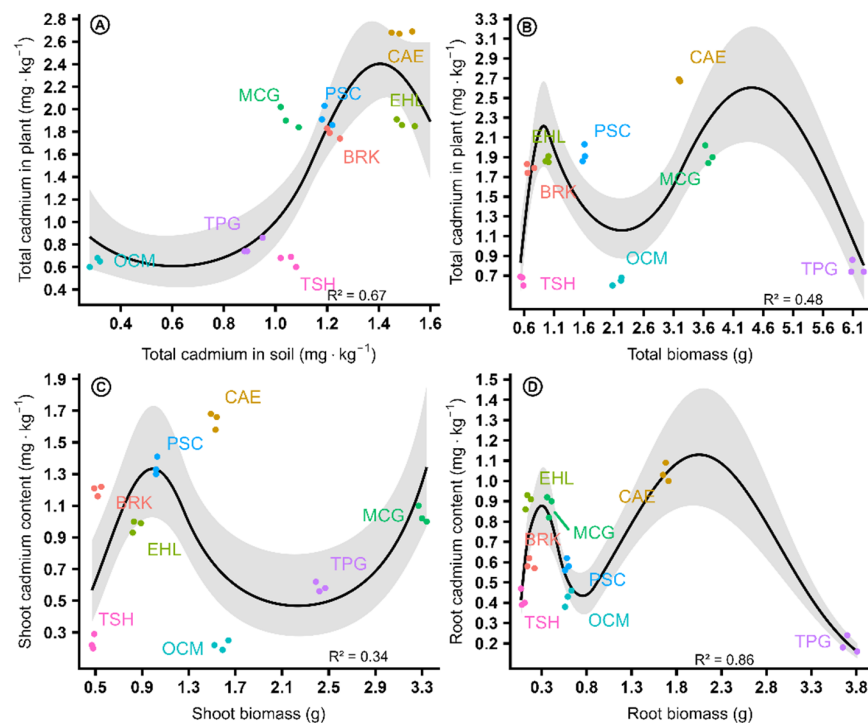
The results indicate a clear relationship between soil Cd content and Cd accumulation in weed species, modeled using generalized linear models with spline functions based on different bases and polynomial degrees. Fig. 1A illustrates the functional relationship between total soil Cd and total Cd in weeds ( $R^2 = 0.67$ ,  $df = 3$ ; polynomial degree = 2). Soils with lower Cd concentrations were associated with *O. campechianum* and *T. paniculatum*, whereas *T. serrulata*; *M. coromandelianum* exhibited intermediate soil Cd levels. In contrast, the highest soil Cd concentrations were recorded in association with *E. heterophylla*, *C. aggregatus*, *B. riparia*, and *P. spiralis*, which also showed intermediate to high levels of Cd accumulation in their tissues. Regarding plant metal concentrations, *C. aggregatus* stood out by exhibiting the highest Cd concentration  $2.68 \text{ mg kg}^{-1}$ .

Fig. 1B depicts the relationship between total biomass and Cd accumulated in plant tissues ( $R^2 = 0.48$ ,  $df = 4$ ; polynomial degree = 2). Species with lower biomass, such as *T. serrulata* and *O. campechianum*, recorded lower Cd contents. *M. coromandelianum* showed intermediate biomass and Cd accumulation, whereas *C. aggregatus* displayed intermediate biomass but the highest Cd accumulation among the evaluated species. In contrast, *T. paniculatum*, despite producing the greatest biomass, exhibited one of the lowest Cd concentrations in plant tissues.

Fig. 1C relates shoot biomass to shoot Cd concentration ( $R^2 = 0.34$ ,  $df = 4$ ; polynomial degree = 2). The species with the lowest shoot biomass, *T. serrulata*, exhibited the lowest Cd concentrations in aerial tissues. Conversely, *E. heterophylla*, *P. spiralis*, despite also presenting low shoot biomass, showed intermediate levels of shoot Cd. *C. aggregatus*, stood out by exhibiting the highest shoot Cd concentration even under low shoot biomass conditions. Likewise, *T. paniculatum*, with intermediate shoot biomass, was among the

species with the lowest Cd concentrations in aerial tissues; whereas *M. coromandelianum* displayed the highest shoot biomass accompanied by intermediate Cd storage.

Fig. 1D illustrates the functional relationship between root biomass and root Cd concentration ( $R^2 = 0.86$ ,  $df = 5$ ; polynomial degree = 2). Species with the lowest root biomass, such as *T. serrulata* and *O. campechianum*, exhibited the lowest levels of root Cd. In contrast, *P. spiralis*, *B. riparia*, *M. coromandelianum* and *E. heterophylla*, despite also presenting low root biomass, showed intermediate root Cd concentrations. *C. aggregatus* displayed intermediate root biomass but the highest root Cd accumulation among the evaluated species. Conversely, *T. paniculatum*, although exhibiting the greatest root biomass, was among the species with the lowest root Cd concentrations.



**Figure 1:** Functional relationships between: **A**) total Cd concentration in soil and total Cd concentration in the plant; **B**) total plant biomass and total Cd concentration in the plant; **C**) aboveground biomass and Cd concentration in the shoot; and **D**) root biomass and Cd concentration in the root. Colored points represent observations for each species. Solid black lines indicate fits from generalized linear models using spline functions, and the shaded areas denote the confidence intervals of the fitted models. The coefficient of determination ( $R^2$ ) reflects the goodness of fit between the evaluated variables. Species abbreviations: PSC: *Pseudelephantopus spiralis* (Less.) Cronquist; OCM: *Ocimum campechianum* Mill; BRK: *Bidens riparia* Kunth; TSH: *Tripogandra serrulata* (Vahl) Handlos; EHL: *Euphorbia heterophylla* L.; TPG: *Talinum paniculatum* (Jacq.) Gaertn; CAE: *Cyperus aggregatus* (Willd.) Endl; MCG: *Malvastrum coromandelianum* (L.) Garcke.

### 3.3 Lead in Weed Species

The weed species exhibited significant differences in both biomass production and Pb concentration ( $p < 0.05$ ) (Table 4). *T. paniculatum* recorded the highest total biomass (6.15 g) and root biomass (3.72 g), while ranking third in shoot biomass (2.43 g). This species was associated with one of the soils that ranked fourth in Pb concentration ( $12.08 \text{ mg kg}^{-1}$ ). In addition, *T. paniculatum* ranked fourth in total Pb accumulation in plant tissues ( $6.06 \text{ mg kg}^{-1}$ ), second in root Pb accumulation ( $3.98 \text{ mg kg}^{-1}$ ), and sixth in



shoot Pb accumulation ( $2.08 \text{ mg kg}^{-1}$ ). In contrast, *A. stachyura* Pax exhibited the highest shoot biomass (4.03 g) and was associated with soils showing the highest Pb concentration ( $14.90 \text{ mg kg}^{-1}$ ), while also recording the highest Pb concentrations in aerial tissues ( $4.15 \text{ mg kg}^{-1}$ ).

*T. serrulata* exhibited the lowest total biomass (0.57 g), root biomass (0.10 g), and shoot biomass (0.48 g); however, its rhizospheric soil recorded the fourth highest Pb concentration ( $12.16 \text{ mg kg}^{-1}$ ). Regarding Pb levels in plant tissues, this species showed the highest total Pb concentration ( $11.19 \text{ mg kg}^{-1}$ ) and root Pb concentration ( $8.02 \text{ mg kg}^{-1}$ ), while ranking fourth in shoot Pb concentration ( $3.17 \text{ mg kg}^{-1}$ ). In contrast, the rhizospheric soil of *C. aggregatus* recorded a Pb concentration of  $13.08 \text{ mg kg}^{-1}$ , ranking second among the evaluated soils. Nevertheless, this species exhibited the lowest total Pb concentration in plant tissues ( $2.59 \text{ mg kg}^{-1}$ ) and the lowest shoot Pb concentration ( $0.56 \text{ mg kg}^{-1}$ ), while also ranking among the species with the lowest root Pb concentrations ( $2.03 \text{ mg kg}^{-1}$ ). Additionally, *C. aggregatus* ranked fourth in both total biomass (3.20 g) and shoot biomass (1.52 g), and third in root biomass (1.68 g).

**Table 4:** Lead concentrations in soil, roots, and shoots, and biomass across weed species: ANOVA, model selection, and estimated marginal means.

| Factors                             | Pb Soil<br>( $\text{mg kg}^{-1}$ ) | Pb Total<br>( $\text{mg kg}^{-1}$ ) | Pb Root<br>( $\text{mg kg}^{-1}$ ) | Pb Shoot<br>( $\text{mg kg}^{-1}$ ) | Total<br>Biomass (g) | Root<br>Biomass (g) | Shoot<br>Biomass (g) |
|-------------------------------------|------------------------------------|-------------------------------------|------------------------------------|-------------------------------------|----------------------|---------------------|----------------------|
| Species                             | 0.00                               | 0.00                                | 0.00                               | 0.00                                | 0.00                 | 0.00                | 0.00                 |
| Model selection and fit             |                                    |                                     |                                    |                                     |                      |                     |                      |
| Distribution                        | lognormal                          | lognormal                           | Gamma                              | Lognormal                           | lognormal            | lognormal           | Gamma                |
| AICc                                | -18.18                             | -9.27                               | -31.06                             | -41.43                              | -39.88               | -57.06              | -62.90               |
| R <sup>2</sup>                      | 0.99                               | 0.99                                | 0.99                               | 0.99                                | 0.99                 | 0.99                | 0.99                 |
| Estimated marginal means by species |                                    |                                     |                                    |                                     |                      |                     |                      |
| ASP                                 | 14.90a                             | 6.70c                               | 2.55d                              | 4.15a                               | 5.75b                | 1.72c               | 4.03a                |
| BRK                                 | 12.44c                             | 5.36e                               | 2.02f                              | 3.33c                               | 0.69g                | 0.18e               | 0.52g                |
| CAE                                 | 13.08b                             | 2.59g                               | 2.03f                              | 0.56g                               | 3.20d                | 1.68c               | 1.52d                |
| EHL                                 | 11.49e                             | 4.55f                               | 2.01f                              | 2.53e                               | 1.00f                | 0.15e               | 0.85f                |
| HLW                                 | 11.25f                             | 5.50e                               | 2.25e                              | 3.26cd                              | 5.48c                | 2.51b               | 2.97b                |
| PSC                                 | 11.18f                             | 6.91b                               | 2.96c                              | 3.95b                               | 1.61e                | 0.58d               | 1.02e                |
| TPG                                 | 12.08d                             | 6.06d                               | 3.98b                              | 2.08f                               | 6.15a                | 3.72a               | 2.43c                |
| TSH                                 | 12.16d                             | 11.19a                              | 8.02a                              | 3.17d                               | 0.57h                | 0.10f               | 0.48h                |

Note: Different letters indicate significant differences among species ( $p < 0.05$ ). PSC: *Pseudelephantopus spiralis* (Less.) Cronquist; BRK: *Bidens riparia* Kunth; TSH: *Tripogandra serrulata* (Vahl) Handlos; EHL: *Euphorbia heterophylla* L.; TPG: *Talinum paniculatum* (Jacq.) Gaertn; HIW: *Hillieria latifolia* (Lam.) H. Walter; CAE: *Cyperus aggregatus* (Willd.) Endl; ASP: *Acalypha stachyura* Pax.

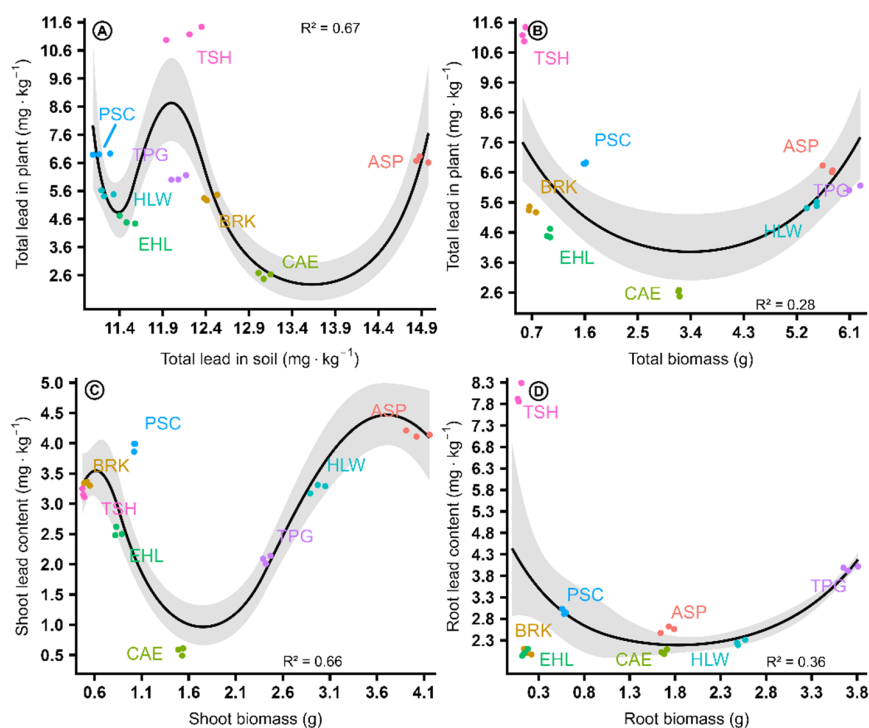
Fig. 2 illustrates the overall relationships between soil Pb content and weed biomass, modeled using generalized linear models with spline functions based on different bases and polynomial degrees. Fig. 2A shows the relationship between soil Pb concentration and Pb accumulation in weed species ( $R^2 = 0.67$ ,  $df = 4$ , degree = 2). At lower soil Pb levels, *E. heterophylla*, *H. latifolia*, *B. riparia* and *T. paniculatum* were recorded and were associated with lower Pb concentrations; however, *T. serrulata*, even within this lower range, exhibited the highest Pb concentration among the species. Soils with intermediate Pb concentrations were represented by *C. aggregatus*, which showed the lowest Pb values, whereas soils with the highest Pb levels were associated with the presence of *A. stachyura*, which exhibited intermediate Pb concentrations.

Fig. 2B describes the relationship between total biomass and Pb concentration in weed species ( $R^2 = 0.28$ ,  $df = 2$ , degree = 2). Species with lower biomass, such as *E. heterophylla*, *B. riparia* and *P. spiralis*, exhibited intermediate Pb concentrations; however, *T. serrulata*, despite belonging to this low-biomass group, recorded the highest Pb. Conversely, *C. aggregatus* showed intermediate biomass levels but the lowest Pb content,

whereas *A. stachyura*, *T. paniculatum*, and *H. latifolia* were characterized by higher biomass and intermediate Pb concentrations. These patterns indicate that metal accumulation is not solely dependent on plant size, but rather on internal processes of uptake, transport, and partitioning associated with species-specific traits and edaphic metal availability, as widely documented in the phytoremediation literature [32].

Fig. 2C shows the relationship between shoot biomass and shoot Pb concentration ( $R^2 = 0.66$ ,  $df = 4$ , degree = 2). Species with lower shoot biomass included *C. aggregatus*, which also recorded the lowest shoot Pb concentration, as well as *E. heterophylla*, *T. serrulata*, and *B. riparia*, which exhibited intermediate shoot Pb concentrations. Notably, *P. spiralis*, despite belonging to this low-shoot-biomass group, showed one of the highest Pb concentrations in aerial tissues. In contrast, *T. paniculatum* and *H. latifolia* displayed intermediate shoot biomass values accompanied by intermediate Pb concentrations, whereas *A. stachyura*, characterized by the highest shoot biomass, ranked among the species with the highest shoot Pb concentration.

Finally, Fig. 2D illustrates the relationship between root biomass and root Pb concentration ( $R^2 = 0.36$ ,  $df = 2$ , degree = 2). Species with lower root biomass, such as *E. heterophylla*, *B. riparia*, and *P. spiralis*, exhibited the lowest root Pb concentrations. Likewise, species with intermediate root biomass values, including *C. aggregatus*, *A. stachyura*, and *H. latifolia*, as well as *T. paniculatum*, which exhibited the highest root biomass, were among the species with the lowest root Pb concentrations. However, *T. serrulata*, despite belonging to this group, showed the highest root Pb concentration.



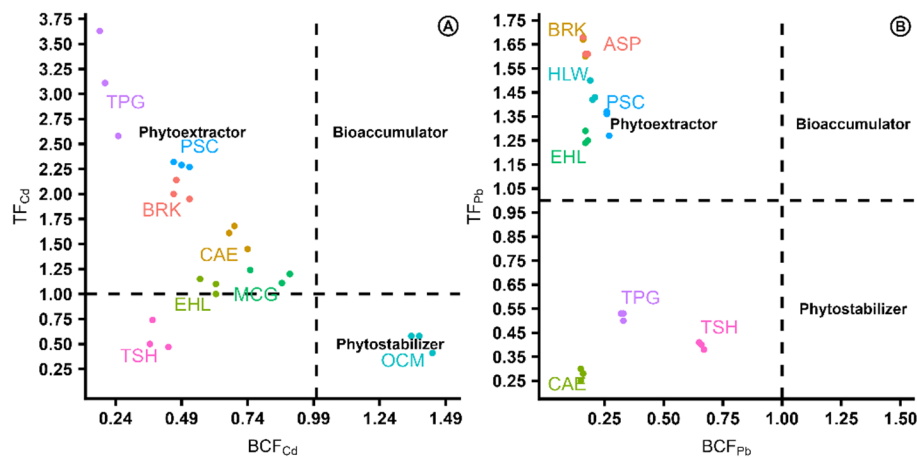
**Figure 2:** Functional relationships between: (A) total Pb concentration in soil and total Pb concentration in the plant; (B) total plant biomass and total Pb concentration in the plant; (C) aboveground biomass and Pb concentration in the shoot; and (D) root biomass and Pb concentration in the root. Colored points represent observations for each species. Solid black lines correspond to generalized linear model fits using spline functions, and the shaded areas represent the model confidence intervals. The coefficient of determination ( $R^2$ ) indicates the goodness of fit between the evaluated variables. Species abbreviations: PSC: *Pseudelephantopus spiralis* (Less.) Cronquist; OCM: *Ocimum campechianum* Mill; BRK: *Bidens riparia* Kunth; TSH: *Tripogandra serrulata* (Vahl) Handlos; EHL: *Euphorbia heterophylla* L.; TPG: *Talinum paniculatum* (Jacq.) Gaertn; CAE: *Cyperus aggregatus* (Willd.) Endl.; MCG: *Malvastrum coromandelianum* (L.) Garcke.

### 3.4 Phytoremediation Potential Based on BCF and TF Factors

Fig. 3 shows the functional classification of the evaluated species according to their bioconcentration factor (BCF) and translocation factor (TF) for Cd and Pb, which allowed the identification of their potential roles as phytoextractors, bioaccumulators, or phytostabilizers.

For Cd (Fig. 3A), the evaluated species were distributed across three of the four quadrants defined by the  $BCF = 1$  and  $TF = 1$  thresholds. Species classified as phytoextractors ( $TF > 1$  and  $BCF < 1$ ) included *T. paniculatum* (TPG), *P. spiralis* (PSC), *C. aggregatus* (CAE), *B. riparia* (BRK), *E. heterophylla* (EHL), and *M. coromandelianum* (MCG). The phytostabilization quadrant ( $BCF > 1$  and  $TF < 1$ ) was represented by *O. campechianum* (OCM), indicating a predominant root retention of the metal and limited internal translocation. In contrast, no species were positioned within the bioaccumulator quadrant ( $BCF > 1$  and  $TF > 1$ ). Finally, *T. serrulata* (TSH) was located in the non-classified quadrant ( $TF < 1$  and  $BCF < 1$ ).

In the case of Pb (Fig. 3B), the distribution pattern of the species reflected a more pronounced differentiation of phytoremediation potential compared with Cd. *B. riparia* (BRK), *A. stachyura* (ASP), *H. latifolia* (HLW), *P. spiralis* (PSC), and *E. heterophylla* (EHL) were positioned within the phytoextraction quadrant ( $BCF < 1$  and  $TF > 1$ ). In contrast, no species were classified within the bioaccumulator ( $BCF > 1$  and  $TF > 1$ ) or phytostabilization ( $BCF > 1$  and  $TF < 1$ ) quadrants, suggesting a limited capacity of the evaluated species to combine high accumulation or effective root retention of Pb under the conditions of this study. Finally, *C. aggregatus* (CAE), *T. paniculatum* (TPG), and *T. serrulata* (TSH) were located in the non-classified quadrant ( $BCF < 1$  and  $TF < 1$ ).



**Figure 3:** Functional classification of the evaluated weed species according to their phytoremediation potential, based on the Bioconcentration Factor (BCF) and the Translocation Factor (TF) for Cd and Pb. **(A)** Distribution of species as a function of BCF and TF for Cd; **(B)** corresponding distribution for Pb. Dashed lines at  $BCF = 1$  and  $TF = 1$  delineate the quadrants associated with phytoextraction, phytostabilization, bioaccumulation, and unclassified categories. Points represent the observed values for each species, and the abbreviations correspond to the evaluated species listed as follows: PSC: *Pseudelephantopus spiralis* (Less.) Cronquist; OCM: *Ocimum campechianum* Mill; BRK: *Bidens riparia* Kunth; TSH: *Tripogandra serrulata* (Vahl) Handlos; EHL: *Euphorbia heterophylla* L.; TPG: *Talinum paniculatum* (Jacq.) Gaertn; CAE: *Cyperus aggregatus* (Willd.) Endl; MCG: *Malvastrum coromandelianum* (L.) Garcke; HLW: *Hillieria latifolia* (Lam.) H. Walter; ASP: *Acalypha stachyura* Pax.

## 4 Discussion

Numerous studies have documented the bioremediation potential of weed species growing in soils contaminated with heavy metals [28,30,56]. In the present study, weed species associated with cacao cultivation were identified, which are naturally adapted to local edaphoclimatic conditions. This represents a clear advantage over introduced species commonly used in phytoremediation programs [32]. The recorded species were distributed across eight botanical families, with Asteraceae, Euphorbiaceae, Cyperaceae, and Malvaceae being the most representative. Species from these families have been widely reported in disturbed environments and in soils contaminated with heavy metals [57]. This floristic composition suggests a plant community exhibiting adaptive strategies linked to physiological resilience under environmental stress conditions.

Several of the identified species exhibit wide spatial and ecological distributions. *P. spiralis* and *B. riparia* show variation in their occurrence across zones, which has been associated with geographic and seasonal conditions [58]. *E. heterophylla* has been reported to grow in heavy metal-contaminated soils without significant impairment of its development, a response that may be linked to its rapid growth rate and invasive behavior [59,60]. *A. stachyura* has been documented across several regions of Peru, including Junín and Pasco, occurring between 100 and 1300 m a.s.l. in humid forest ecosystems [61]. Species of the genus *Ocimum* have been reported to possess the ability to block key ion transport channels involved in root-to-shoot translocation processes [62]. *T. serrulata* is a short-lived annual species widely distributed in warm and temperate regions, ranging from sea level up to 1600 m a.s.l. [63,64]. *T. paniculatum*, broadly distributed throughout South America between 0 and 2000 m a.s.l., is characteristic of disturbed environments and can reach heights of up to 60 cm [65]. *H. latifolia* is a perennial herb commonly found in South America and tropical Africa, with stems that may reach up to 2 m in height [66]. *C. aggregatus* shows high adaptability to wet environments and has demonstrated the ability to establish in soils contaminated with hydrocarbons [67,68]. Finally, *Malvastrum coromandelianum* (L.) Garcke is frequently found in cultivated fields and disturbed areas and exhibits broad tolerance to varying light conditions [69].

Regarding biomass production, significant differences were observed between aboveground and belowground organs, with *T. paniculatum* standing out for its highest total biomass (6.15 g) (Table 3). *A. stachyura* and *M. coromandelianum* exhibited the greatest aboveground biomass (4.03 g and 3.30 g, respectively), with *A. stachyura* showing the highest Pb concentration in this compartment (4.15 mg kg<sup>-1</sup>). Similarly, *T. paniculatum* and *H. latifolia* were characterized by high root biomass (3.72 g and 2.51 g, respectively), with *T. paniculatum* displaying one of the highest Pb concentrations in roots (3.98 mg kg<sup>-1</sup>). *C. aggregatus* with an intermediate biomass weight, showed the highest Cd level in the whole plant (2.68 mg kg<sup>-1</sup>). Biomass allocation among plant organs is governed by genotypic traits modulated by plant-environment interactions and depends on factors such as leaf area, climate, nutrient availability, and assimilate fluxes [70]. In species with phytoremediation potential, biomass production is as relevant as the ability to tolerate or accumulate heavy metals [71,72], since hyperaccumulator species such as *Thlaspi caerulescens* and *Arabidopsis halleri* present practical limitations due to their low biomass production [27]. In contrast, cacao-associated weed species such as *Cissus verticillata* have shown operational advantages owing to their higher biomass in soils with elevated Cd concentrations [73].

Phytoremediation potential was further assessed using the bioconcentration factor (BCF) and the translocation factor (TF), which are widely applied indicators for classifying the functional behavior of plants in response to heavy metal contamination [54,74]. According to Mellem et al. [51] and Hidayati et al. [32], BCF and TF values greater than 1 indicate a bioaccumulative behavior, whereas a BCF value

exceeding 2 reflects high accumulation efficiency. In addition, a BCF > 1 combined with a TF < 1 is associated with phytostabilization, while a BCF < 1 and TF > 1 suggests phytoextraction potential.

In this study, significant differences in BCF and TF values for Cd and Pb were observed among the evaluated species (Fig. 3), revealing behaviors consistent with phytostabilization and phytoextraction strategies. *O. campechianum* exhibited a pattern compatible with Cd phytostabilization (BCF < 1, TF > 1), characterized by accumulation in roots and limited translocation to aerial tissues, a mechanism that contributes to metal immobilization in the rhizosphere [75]. Although no previous studies have directly evaluated this species for phytoremediation purposes, related evidence has been reported for *O. basilicum*, which has been described as a Cd bioaccumulator in roots while preserving essential oil quality [76,77]. For Pb, none of the evaluated species met the established criteria for phytostabilization. *C. aggregatus*, *T. serrulata*, and *T. paniculatum* did not fully conform to the classical functional categories. However, *T. serrulata* has been reported to exhibit enhanced root Pb accumulation, suggesting an exclusion behavior [78], which may be useful for metal stabilization and reducing trophic transfer. In the case of *C. aggregatus* although direct evidence is lacking, related species such as *C. elegans* have shown tolerance to Cd and enhanced root growth following inoculation with rhizobacteria [79].

*P. spiralis*, *B. riparia*, *E. heterophylla*, *M. coromandelianum*, *C. aggregatus*, and *Talinum paniculatum* exhibited patterns compatible with phytoextraction potential for Cd. Whereas for Pb, *P. spiralis*, *B. riparia*, *E. heterophylla*, *H. latifolia*, and *A. stachyura* were the most prominent species. Nevertheless, phytoremediation effectiveness also depends on additional attributes such as vigorous growth, biomass production, root system architecture, metal tolerance, and agronomic management [80,81], as well as metal bioavailability in the rhizosphere and its translocation to aboveground biomass [82]. In this study, the soil exhibited a pH of 6.3, a slightly acidic range that favors Cd availability and plant uptake [83], which may partly explain the observed responses. Likewise, the organic matter content (4.1%) plays a key role in metal bioavailability, as it modulates complexation and adsorption processes that can reduce the fraction available for plant uptake [84]. Therefore, the BCF and TF values obtained should be interpreted within the specific edaphic conditions of the study site, as they may vary considerable across cocoa-growing with different physicochemical characteristics, particularly as a function of soil pH and organic matter content.

The behavior of *P. spiralis* and *B. riparia* (Asteraceae) is particularly noteworthy, as both species are widely documented for their phytoextraction capacity [85–88]. The genus *Bidens* has demonstrated accumulation potential for both Cd and Pb [89–93], in addition to contributing to improvements in soil structure and fertility. *P. spiralis* has been classified as a Cd hyperaccumulator in soils with pH values between 6 and 7, possibly due to the release of root exudates that acidify the rhizosphere and enhance metal bioavailability [73]. *E. heterophylla* exhibits differential affinity for metals in the order Cd > Hg > Pb > As [59], whereas *A. stachyura* has been identified as a Pb accumulator, with antioxidant mechanisms that mitigate metal toxicity [94,95]. In contrast, *M. coromandelianum* has been reported as a phytostabilizer in industrial soils contaminated with Pb and Cu [56], a behavior that differs from the patterns observed in the present study. Finally, *T. paniculatum* stood out by exhibiting the highest TF value for Cd, consistent with previous reports highlighting its rhizofiltration potential, tolerance to edaphic stress, and capacity for the degradation of organic contaminants [96,97].

Similarly, it is important to emphasize that the practical implementation of these species in association with cocoa cultivation should be evaluated through controlled experimental trials that integrate both remediation efficiency and ecological and economic impact. The development of specific management protocols will be essential to ensure that environmental benefits outweigh potential risks and that the strategy remains viable in terms of productive sustainability. Although these weed species may pose certain



challenges, previous studies have reported favorable outcomes. For instance, interspecific competition (i.e., the coexistence between weeds and cocoa under regulated management) has been shown to improve soil structure and enhance moisture retention [28]. Additionally, the propagation capacity of weeds can be harnessed as ground cover to reduce soil erosion [30], while metal-enriched biomass may be repurposed for biochar production or composting [20].

Overall, this study enabled the identification of weed species associated with cacao cultivation that exhibit traits compatible with potential phytoextraction or phytostabilization performance. Although the present results do not constitute direct functional evidence of remediation, they expand current knowledge of spontaneous vegetation in cacao-based systems and provide a technical foundation for the design of controlled experiments aimed at quantifying their actual contribution to the mitigation of heavy metals in agricultural soils. In this context, the findings strengthen the prospect of biologically based solutions applicable to one of the major challenges facing the cacao sector.

## 5 Limitations

Although these results allow the identification of promising patterns of Cd and Pb accumulation and translocation in weed species associated with cacao cultivation, their interpretation should consider an inherent limitation of the study design. While the research initially involved the assessment of seven cacao-producing sites, the identification and detailed analysis of species were ultimately focused on the site exhibiting the highest Cd concentration. This approach is consistent with the explanatory nature of the study, which aimed to maximize the detection of weed species with potential accumulation rate. Targeted sampling strategies in highly contaminated sites have been employed in previous phytoremediation prospecting studies [28,30,56], as they enable the observation of functional responses under real stress conditions. The findings are not intended to be generalizable across all edaphoclimatic conditions of the region; rather, they provide a technical basis for the development of future studies incorporating experimental designs that include multiple sites and management. Therefore, the observed patterns should be interpreted with caution and validated in subsequent multisite and controlled studies to assess their consistency and applicability across different cacao production systems. BCF and TF values obtained should be interpreted within the specific edaphic conditions of the study site, as these indicators may vary considerably in cacao soils with different physicochemical properties, particularly as a function of soil pH and organic matter content. This underscores the importance of subsequent controlled trials to demonstrate the removal of heavy metals under those conditions. In this context, it is essential to develop experimental studies that quantify the removal of Cd and Pb from soils over time and, across different plant growth stages, as well as to evaluate extraction or stabilization efficiency as a function of metal bioavailability and edaphic conditions. This approach will enable validation of the patterns inferred from bioconcentration and translocation factors and support their application in sustainable strategies for the management of contaminated soils.

## 6 Conclusions

The present study enabled the identification of spontaneous weed species associated with cacao cultivation that exhibit contrasting patterns of biomass production, bioconcentration, and translocation of Cd and Pb, revealing behaviors consistent with potential phytoextraction and phytostabilization strategies. The integrated assessment of plant biomass production together with bioconcentration factor (BCF) and translocation factor (TF) proved effective for the functional discrimination of the evaluated species under field conditions. Among the analyzed weeds, *P. spiralis*, *B. riparia*, *E. heterophylla*, *M. coromandelianum*,

*C. aggregatus* and *T. paniculatum* exhibited patterns compatible with potential Cd phytoextraction, whereas *P. spiralis*, *B. riparia*, *E. heterophylla*, *H. latifolia* and *A. stachyura* showed similar responses with respect to Pb. In contrast, *O. campechianum* displayed a behavior consistent with Cd phytostabilization, characterized by metal retention in the root system and limited translocation to aerial tissues. Biomass production emerged as a key component for interpreting phytoremediation potential, complementing the information provided by BCF and TF indices. In this context, species that combine tolerance to heavy metals with higher biomass production appear viable candidates, for future experimental evaluations. Although the results obtained do not constitute functional evidence of soil remediation, they provide relevant insights into the behavior of native weed species under the edaphoclimatic conditions of cacao agroecosystems. These findings expand current knowledge on spontaneous vegetation in cacao-based systems and establish a technical basis for the design of controlled trials aimed at evaluating the actual efficiency of these species in heavy-metal management and mitigation strategies in agricultural soils.

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**Availability of Data and Materials:** The data that support the findings of this study are available from the Corresponding Author, [Lorena E. Romero-Chávez], upon reasonable request.

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## References

1. Abd Rahim FA, Abdul Keyon AS, Kamaruddin AF. Assessment of tin and lead in commercial canned foods and associated health risk indicators. *J Food Compos Anal.* 2025;148:108400. [[CrossRef](#)].
2. Rodríguez Albarreín HS, Darghan Contreras AE, Henao MC. Spatial regression modeling of soils with high cadmium content in a cocoa producing area of Central Colombia. *Geoderma Reg.* 2019;16:e00214. [[CrossRef](#)].
3. Scaccabarozzi D, Castillo L, Aromatizi A, Milne L, Bullon Castillo A, Muñoz-Rojas M. Soil, site, and management factors affecting cadmium concentrations in cacao-growing soils. *Agronomy.* 2020;10(6):806. [[CrossRef](#)].
4. Iqbal M, Iqbal N, Ahmad Bhatti I, Ahmad N, Zahid M. Response surface methodology application in optimization of cadmium adsorption by shoe waste: A good option of waste mitigation by waste. *Ecol Eng.* 2016;88:265–75. [[CrossRef](#)].

5. Rafique N, Tariq SR. Distribution and source apportionment studies of heavy metals in soil of cotton/wheat fields. *Environ Monit Assess.* 2016;188(5):309. [[CrossRef](#)].
6. Arévalo-Gardini E, Arévalo-Hernández CO, Baligar VC, He ZL. Heavy metal accumulation in leaves and beans of cacao (*Theobroma cacao* L.) in major cacao growing regions in Peru. *Sci Total Environ.* 2017;605–606:792–800. [[CrossRef](#)].
7. Florida Rofner N. Revisión sobre límites máximos de cadmio en cacao (*Theobroma cacao* L.). *Granja Rev Cienc Vida.* 2021;34(2):117–30. [In Spanish]. [[CrossRef](#)].
8. Oliva M, Rubio K, Epquin M, Marlo G, Leiva S. Cadmium uptake in native cacao trees in agricultural lands of bagua, Peru. *Agronomy.* 2020;10(10):1551. [[CrossRef](#)].
9. Decreto Supremo N° 011-2017-MINAM Aprueban Estándares de Calidad Ambiental (ECA) para Suelo. Lima, Perú: Diario El Peruano; 2017. [In Spanish].
10. Comisión Europea. REGLAMENTO (UE) N° 488/2014 DE LA COMISIÓN de 12 de mayo de 2014 que modifica el Reglamento (CE) n° 1881/2006 por lo que respecta al contenido máximo de cadmio en los productos alimenticios. Luxembourg City, Luxembourg: Diario Oficial de la Unión Europea; 2014. [In Spanish].
11. Codex Alimentarius. Informe de la 14.<sup>a</sup> reunión del comité del codex sobre contaminantes de los alimentos. Geneva, Switzerland: Organización Mundial de la Salud; 2021. [In Spanish].
12. Thomas E, Atkinson R, Zavaleta D, Rodriguez C, Lastra S, Yovera F, et al. The distribution of cadmium in soil and cacao beans in Peru. *Sci Total Environ.* 2023;881:163372. [[CrossRef](#)].
13. Bertoldi D, Barbero A, Camin F, Caligiani A, Larcher R. Multielemental fingerprinting and geographic traceability of *Theobroma cacao* beans and cocoa products. *Food Control.* 2016;65:46–53. [[CrossRef](#)].
14. Argüello D, Chavez E, Lauryssen F, Vanderschueren R, Smolders E, Montalvo D. Soil properties and agronomic factors affecting cadmium concentrations in cacao beans: A nationwide survey in Ecuador. *Sci Total Environ.* 2019;649:120–7. [[CrossRef](#)].
15. Guzmán JH, Chire Fajardo GC. Evaluación de la cadena de valor del cacao (*Theobroma cacao* l.) peruano. Enfoque UTE. 2019;10(1):97–116. [In Spanish]. [[CrossRef](#)].
16. Yang W, Qi L, Gusiati M. Editorial: Remediation and health risks of heavy metal contaminated soils. *Front Environ Sci.* 2024;12:1501443. [[CrossRef](#)].
17. Cui Y, Wang X, Wang X, Zhang X, Fang L. Evaluation methods of heavy metal pollution in soils based on enzyme activities: A review. *Soil Ecol Lett.* 2021;3(3):169–77. [[CrossRef](#)].
18. Nkansah MA, Korankye M, Darko G, Dodd M. Heavy metal content and potential health risk of geophagic white clay from the Kumasi Metropolis in Ghana. *Toxicol Rep.* 2016;3:644–51. [[CrossRef](#)].
19. Sun W, Xiong Z, Chu L, Li W, Soares MA, White JF, et al. Bacterial communities of three plant species from Pb-Zn contaminated sites and plant-growth promotional benefits of endophytic *Microbacterium* sp. (strain BXGe<sub>71</sub>). *J Hazard Mater.* 2019;370:225–31. [[CrossRef](#)].
20. Shah V, Daverey A. Phytoremediation: A multidisciplinary approach to clean up heavy metal contaminated soil. *Environ Technol Innov.* 2020;18:100774. [[CrossRef](#)].
21. Clabeaux BL, Navarro DAG, Aga DS, Bisson MA. Cd tolerance and accumulation in the aquatic macrophyte, *Chara australis*: Potential use for charophytes in phytoremediation. *Environ Sci Technol.* 2011;45(12):5332–8. [[CrossRef](#)].
22. Galvis DA, Jaimes-Suárez YY, Rojas Molina J, Ruiz R, Carvalho FEL. Cadmium up taking and allocation in wood species associated to cacao agroforestry systems and its potential role for phytoextraction. *Plants.* 2023;12(16):2930. [[CrossRef](#)].
23. Carvalho FEL, Montenegro AC, Escobar-Pachajoa LD, Rojas-Molina J, Camacho-Diaz JE, Rengifo-Estrada GA. Phytoextraction and Cd allocation to the stem of woody species used in cacao agroforestry. *Plants.* 2025;14(7):1101. [[CrossRef](#)].
24. Kayser A, Wenger K, Keller A, Attinger W, Felix HR, Gupta SK, et al. Enhancement of phytoextraction of Zn, Cd, and Cu from calcareous soil: The use of NTA and sulfur amendments. *Environ Sci Technol.* 2000;34(9):1778–83. [[CrossRef](#)].
25. Herzig R, Nehnevajova E, Pfister C, Schwitzguebel JP, Ricci A, Keller C. Feasibility of labile Zn phytoextraction using enhanced tobacco and sunflower: Results of five- and one-year field-scale experiments in Switzerland. *Int J Phytoremediat.* 2014;16(7–12):735–54. [[CrossRef](#)].

26. Wani KA, Sofi ZM, Malik JA, Wani JA. Phytoremediation of heavy metals using salix (willows). In: Bhat RA, Hakeem KR, Dervash MA, editors. Bioremediation and biotechnology. Vol. 2. Cham, Switzerland: Springer International Publishing; 2020. p. 161–74. [CrossRef].
27. Mahar A, Wang P, Ali A, Awasthi MK, Lahori AH, Wang Q, et al. Challenges and opportunities in the phytoremediation of heavy metals contaminated soils: A review. *Ecotoxicol Environ Saf.* 2016;126:111–21. [CrossRef].
28. Wang J, Xiong Y, Zhang J, Lu X, Wei G. Naturally selected dominant weeds as heavy metal accumulators and excluders assisted by rhizosphere bacteria in a mining area. *Chemosphere.* 2020;243:125365. [CrossRef].
29. Wei S, Zhou Q, Wang X. Identification of weed plants excluding the uptake of heavy metals. *Environ Int.* 2005;31(6):829–34. [CrossRef].
30. Singh N, Kaur M, Katnoria JK. Analysis on bioaccumulation of metals in aquatic environment of Beas River Basin: A case study from Kanjli wetland. *Geohealth.* 2017;1(3):93–105. [CrossRef].
31. Ramadan WF, Balah MA. The use of some weeds type in the disposal of heavy metals in contaminated soil. *J Saudi Soc Agric Sci.* 2022;21(5):289–95. [CrossRef].
32. Hidayati N, Rini DS. Assessment of plants as lead and cadmium accumulators for phytoremediation of contaminated rice fields. *Biodiversitas.* 2020;21(5):1928–34. [CrossRef].
33. Chinmayee MD, Mahesh B, Pradesh S, Mini I, Swapna TS. The assessment of phytoremediation potential of invasive weed *Amaranthus spinosus* L. *Appl Biochem Biotechnol.* 2012;167(6):1550–9. [CrossRef].
34. Azab E, Hegazy AK. Monitoring the efficiency of *Rhazya stricta* L. plants in phytoremediation of heavy metal-contaminated soil. *Plants.* 2020;9(9):1057. [CrossRef].
35. Instituto Nacional de Recursos Naturales. Mapa ecologico del Perú. Lima, Perú: Ministerio de Agricultura; 1995. [In Spanish].
36. SENAMHI—Mapa Climatico del Peru [Internet]. [cited 2025 Mar 11]. Available from: <https://www.senamhi.gob.pe/?p=mapa-climatico-del-peru>. [In Spanish].
37. Havlin JL, Tisdale SL, Nelson WD, Beaton JD. Soil fertility and fertilizers: An introduction to nutrient management. 6th ed. Vol. 8. Chennai, India: Pearson Education India; 2016.
38. ISO 11464. Soil quality—Pretreatment of samples for physico-chemical analysis. Geneva, Switzerland: ISO (International Organization for Standardization); 2006. [cited 2026 Jan 15] Available from: <https://cdn.standards.iteh.ai/samples/37718/fb0dabb3c1374ae4afad5966da0a0125/ISO-11464-2006.pdf>
39. Norma Oficial Mexicana NOM-021-RECNAT-2000. Norma Oficial Mexicana que establece las especificaciones de fertilidad, salinidad y clasificación de suelos. Estudios, muestreo y análisis. México City, México: Diario Oficial de la Federación; 2022 [cited 2024 Aug 13]. Available from: <https://faolex.fao.org/docs/pdf/mex50674.pdf>. [In Spanish].
40. USEPA. Method 9045D. Soil and waste pH 2004. Washington, DC, USA: USEPA; 2004 [cited 2024 Aug 13]. Available from: <https://www.epa.gov/sites/default/files/2015-12/documents/9045d.pdf>.
41. ISO 10694:1995. Soil quality—Determination of organic and total carbon after dry combustion (elementary analysis). Geneva, Switzerland: ISO (International Organization for Standardization); 1995.
42. ISO 11261. Soil quality—Determination of total nitrogen—Modified Kjeldahl method. Geneva, Switzerland: ISO (International Organization for Standardization); 1995.
43. Bazan R. Manual para el análisis químico de suelos, plantas y aguas. Lima, Perú: Universidad Nacional Agraria La Molina; 1996. [In Spanish].
44. U.S. EPA. EPA Method 3050B: Acid digestion of sediments, sludges, and soils [data and tools] [Internet]. [cited 2025 Jun 25]. Available from: <https://www.epa.gov/esam/epa-method-3050b-acid-digestion-sediments-sludges-and-soils>.
45. Argüello D, Dekeyrel J, Chavez E, Smolders E. Gypsum application lowers cadmium uptake in cacao in soils with high cation exchange capacity only: A soil chemical analysis. *Eur J Soil Sci.* 2022;73(2):e13230. [CrossRef].
46. Clemente Huachen JP, Medina Contreras J, Laura Pfuño JD, Pariona Aguilar LÁ, Gutierrez Vilchez PP. Fitorremediación en suelos contaminados con Cd usando girasol (*Helianthus annuus* L. var. Sunbright). *Acta Agron.* 2021;70(2):163–70. [In Spanish]. [CrossRef].
47. Lot A, Chiang F. Manual de herbario, administración y manejo de colecciones técnicas de recolección y preparación de ejemplares botánicos. México City, México: Instituto de Biología; 1986. [In Spanish].

48. Alanís E, Mora A, Marroquín J. Muestreo ecológico de la vegetación. San Nicolás de los Garza, México: Universidad Autónoma de Nuevo León; 2020. [In Spanish].
49. The Angiosperm Phylogeny Group. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. Bot J Linn Soc. 2016;181(1):1–20. [[CrossRef](#)].
50. Nayak AK, Raja R, Rao KS, Shukla AK, Mohanty S, Shahid M, et al. Effect of fly ash application on soil microbial response and heavy metal accumulation in soil and rice plant. Ecotoxicol Environ Saf. 2015;114:257–62. [[CrossRef](#)].
51. Mellem JJ. Bioaccumulation of Cr, Hg, As, Pb, Cu and Ni with the ability for hyperaccumulation by *Amaranthus dubius*. Afr J Agric Res. 2012;7(4):591–6. [[CrossRef](#)].
52. Zhao X, Joo JC, Lee JK, Kim JY. Mathematical estimation of heavy metal accumulations in *Helianthus annuus* L. with a sigmoid heavy metal uptake model. Chemosphere. 2019;220:965–73. [[CrossRef](#)].
53. Yan A, Wang Y, Tan SN, Mohd Yusof ML, Ghosh S, Chen Z. Phytoremediation: A promising approach for revegetation of heavy metal-polluted land. Front Plant Sci. 2020;11:359. [[CrossRef](#)].
54. Aziz RA, Yiwen M, Saleh M, Salleh MN, Gopinath SCB, Giap SGE, et al. Bioaccumulation and translocation of heavy metals in paddy (*Oryza sativa* L.) and soil in different land use practices. Sustainability. 2023;15(18):13426. [[CrossRef](#)].
55. Ramírez A, García G, Werner O, Navarro-Pedreño J, Ros RM. Implications of the phytoremediation of heavy metal contamination of soils and wild plants in the industrial area of Haina, Dominican Republic. Sustainability. 2021;13(3):1403. [[CrossRef](#)].
56. Nazir A, Malik RN, Ajaib M, Khan N, Siddiqui MF. Hyperaccumulators of heavy metals of industrial areas of Islamabad and Rawalpindi. Pak J Bot. 2011;43(4):1925–33.
57. Chan-Quijano JG, Cach-Pérez MJ, Cach-Pérez MJ, López-Mejía M, López-Mejía M. Especies vegetales con uso potencial en la remediación de zonas contaminadas en México. Rev For Del Perú. 2021;36(1):22. [In Spanish]. [[CrossRef](#)].
58. Nguyen DTC, Le HTN, Nguyen TT, Nguyen TTT, Liew RK, Bach LG, et al. Engineering conversion of Asteraceae plants into biochars for exploring potential applications: A review. Sci Total Environ. 2021;797:149195. [[CrossRef](#)].
59. John Onyinye Anarado C, Ebere Anarado C, Chisom Nwajide F, Obiora A, Dan Chukwuma V, Lilian Anarado I. Effect of fertilizer application on phytoremediating potentials of *Euphorbia heterophylla*, *Axonopus compressus*, *Emilia coccinea* and *Scoparia dulcis* against  $Hg^{2+}$ ,  $Pb^{2+}$ ,  $Cd^{2+}$  and  $As^{3+}$ . Orient J Chem. 2020;36(3):474–80. [[CrossRef](#)].
60. Trezzi MM, Alcántara-de la Cruz R, Rojano-Delgado AM, Alcántara E, De Bortoli Pagnoncelli F Jr, Viecelli M, et al. Influence of temperature on the retention, absorption and translocation of fomesafen and imazamox in *Euphorbia heterophylla*. Pestic Biochem Physiol. 2021;173:104794. [[CrossRef](#)].
61. Cardiel JM, Muñoz P. Synopsis of *Acalypha* (Euphorbiaceae) of continental Ecuador. PhytoKeys. 2012;17:1–17. [[CrossRef](#)].
62. Gajić G, Djurdjević L, Kostić O, Jarić S, Mitrović M, Pavlović P. Ecological potential of plants for phytoremediation and ecorestoration of fly ash deposits and mine wastes. Front Environ Sci. 2018;6:124. [[CrossRef](#)].
63. Espejo-Serna A, López-Ferrari AR, Ceja-Romero J. Familia commelinaceae. In: Flora del bajío y de reg adyacentes. Xalapa, México: Instituto de Ecología A. C.; 2009. [[CrossRef](#)].
64. De Oliveira Pellegrini MO, Aona-Pinheiro LYS, Forzza RC. Taxonomy and conservation status of *Tripogandra warmingiana* (Seub.) Handlos (Commelinaceae), a previously obscure taxon from Brazil. Phytotaxa. 2013;91(2):39–49. [[CrossRef](#)].
65. Dos Reis LFC, Cerdeira CD, De Paula BF, da Silva JJ, Coelho LFL, Silva MA, et al. Chemical characterization and evaluation of antibacterial, antifungal, antimycobacterial, and cytotoxic activities of *Talinum paniculatum*. Rev Inst Med Trop Sao Paulo. 2015;57(5):397–405. [[CrossRef](#)].
66. Odewo SA, Nwankwo OE, Adeniyi IM, Odozie EC. Comparative studies of two medicinal plants: *Petiveria alliacea* L. and *Hillieria latifolia* (Lam.) H. Walter (Petiveriaceae) based on foliar anatomy. Plants Environ. 2020;2(2):54–8. [[CrossRef](#)].
67. Merkl N, Schultze-Kraft R, Infante C. Assessment of tropical grasses and legumes for phytoremediation of petroleum-contaminated soils. Water Air Soil Pollut. 2005;165(1):195–209. [[CrossRef](#)].
68. Gómez-Laurito J. Las ciperáceas (Cyperaceae) de la Estación Biológica La Selva, Costa Rica. Rev Biol Trop. 2009;57(1):93–110. [In Spanish].



69. Galíndez G, Lopez-Spahr D, Gomez CA, Pastrana-Ignes V, Diaz R, Bertuzzi T, et al. Estructura morfo-anatómica y salida de dormición física de las semillas de *Malvastrum coromandelianum* ssp. *coromandelianum* (Malvaceae). Bol Soc Argent Bot. 2019;54(4):509–17. [In Spanish]. [[CrossRef](#)].
70. Van Heemst HDJ. The distribution of dry matter during growth of a potato crop. Potato Res. 1986;29(1):55–66. [[CrossRef](#)].
71. Salt DE, Blaylock M, Kumar NP, Dushenkov V, Ensley BD, Chet I, et al. Phytoremediation: A novel strategy for the removal of toxic metals from the environment using plants. Biotechnology. 1995;13(5):468–74. [[CrossRef](#)].
72. Kargar M, Clark OG, Hendershot WH, Jutras P, Prasher SO. Immobilization of trace metals in contaminated urban soil amended with compost and biochar. Water Air Soil Pollut. 2015;226(6):191. [[CrossRef](#)].
73. Carrillo Zenteno MD, Orosco Maldonado MP, Peña Jordan AK, Martínez Buñay LM, Peña Salazar KE, García-Orellana Y. Evaluación de arvenses como fitorremediadoras de suelos cacaoteros contaminados con cadmio. Rev Int De Contam Ambient. 2025;41. [In Spanish]. [[CrossRef](#)].
74. Suman J, Uhlik O, Viktorova J, Macek T. Phytoextraction of heavy metals: A promising tool for clean-up of polluted environment? Front Plant Sci. 2018;9:1476. [[CrossRef](#)].
75. Shackira AM, Puthur JT. Phytostabilization of heavy metals: Understanding of principles and practices. In: Srivastava S, Srivastava AK, Suprasanna P, editors. Plant-metal interactions. Cham, Switzerland: Springer International Publishing; 2019. p. 263–82. [[CrossRef](#)].
76. Dinu C, Vasile GG, Buleandra M, Popa DE, Gheorghe S, Ungureanu EM. Translocation and accumulation of heavy metals in *Ocimum basilicum* L. plants grown in a mining-contaminated soil. J Soils Sediments. 2020;20(4):2141–54. [[CrossRef](#)].
77. Alamo-Nole L, Estrella-Martinez B. Phytoremediation of CdS/Te quantum dots by *Ocimum basilicum* in the presence of EDTA. J Environ Health Sci Eng. 2022;20(2):827–34. [[CrossRef](#)].
78. Carusso S, Rodriguez N, Juárez ÁB, El Kassisse Y, Rodriguez Salemi V, de Cabo LI. Phytoremediation potential of *Tripogandra diuretica* for chromium and lead contaminated soils. Int J Environ Sci Technol. 2025;22(3):1313–26. [[CrossRef](#)].
79. Labra-Cardón D, Guerrero-Zúñiga L, Rodríguez-Tovar A, Montes-Villafán S, Pérez-Jiménez S, Rodríguez-Dorantes A. Respuesta de crecimiento y tolerancia a metales pesados de *Cyperus elegans* y *Echinochloa polystachya* inoculadas con una rizobacteria aislada de un suelo contaminado con hidrocarburos derivados del petróleo. Rev Int Contam Ambient. 2012;28(1):7–16. [In Spanish].
80. Robinson BH, Anderson CWN, Dickinson NM. Phytoextraction: Where's the action? J Geochem Explor. 2015;151:34–40. [[CrossRef](#)].
81. Sheoran V, Sheoran AS, Poonia P. Factors affecting phytoextraction: A review. Pedosphere. 2016;26(2):148–66. [[CrossRef](#)].
82. McGrath SP, Zhao FJ. Phytoextraction of metals and metalloids from contaminated soils. Curr Opin Biotechnol. 2003;14(3):277–82. [[CrossRef](#)].
83. Kicińska A, Pomykała R, Izquierdo-Diaz M. Changes in soil pH and mobility of heavy metals in contaminated soils. Eur J Soil Sci. 2022;73(1):e13203. [[CrossRef](#)].
84. Xin J. Enhancing soil health to minimize cadmium accumulation in agro-products: The role of microorganisms, organic matter, and nutrients. Environ Pollut. 2024;348:123890. [[CrossRef](#)].
85. Rodrigo Tintaya DJ. Estimación de la capacidad fitorremediadora del “girasol” *Helianthus annuus* mediante la incorporación de enmiendas para suelos contaminados por metales pesados (Plomo, Cromo) de industrias metalmeccánicas. Ciencia. 2018;4(1):23–8. [In Spanish]. [[CrossRef](#)].
86. Dridi N, Bouslimi H, Caçador I, Sleimi N. Lead tolerance, accumulation and translocation in two Asteraceae plants: *Limbaria crithmoides* and *Helianthus annuus*. S Afr N J Bot. 2022;150:986–96. [[CrossRef](#)].
87. Niu Z, Li X, Mahamood M. Accumulation potential cadmium and lead by sunflower (*Helianthus annuus* L.) under citric and glutaric acid-assisted phytoextraction. Int J Environ Res Public Health. 2023;20(5):4107. [[CrossRef](#)].
88. Waseem M, Khilji SA, Tariq S, Jamal A, Alomrani SO, Javed T. Phytoremediation of heavy metals from industrially contaminated soil using sunflower (*Helianthus annuus* L.) by inoculation of two indigenous bacteria. Plant Stress. 2024;11:100297. [[CrossRef](#)].
89. Dou X, Dai H, Skuza L, Wei S. *Bidens pilosa* L. hyperaccumulating Cd with different species in soil and the role of EDTA on the hyperaccumulation. Environ Sci Pollut Res Int. 2019;26(25):25668–75. [[CrossRef](#)].

90. Bobadilla Roman WJ, Valderrama De La Cruz JA, Lujan Rojas JM, Llaque Fernández GI, Valderrama Puscan MW. Phytoremediation of heavy metals in soil. A systematic review of the literature between the years 2012–2022. In: Proceedings of the 2nd LACCEI International Multiconference on Entrepreneurship, Innovation and Regional Development (LEIRD 2022); 2022 Dec 6–7; Virtual. [[CrossRef](#)].
91. Yu F, Tang S, Shi X, Liang X, Liu K, Huang Y, et al. Phytoextraction of metal(loid)s from contaminated soils by six plant species: A field study. *Sci Total Environ.* 2022;804:150282. [[CrossRef](#)].
92. Chico-Ruiz J, Cerna-Rebaza L, Gonzalez Llontop LF, del Rocío Chotón Calvo M, Saldaña Miranda MY, Rodríguez Rosales CS. El exceso de cobre inhibe el crecimiento de *Bidens pilosa* en condiciones de laboratorio. *Revistaalfa.* 2022;6(16):166–77. [In Spanish]. [[CrossRef](#)].
93. Shakeel T, Shah GM, Zeb BS, Gul I, Bibi S, Hussain Z, et al. Phytoremediation potential and vegetation assessment of plant species growing on multi-metals contaminated coal mining site. *Environ Res Commun.* 2024;6(5):055006. [[CrossRef](#)].
94. Venkateshappa SM, Sreenath KP. Potential medicinal plants of Lamiaceae. *Am Int J Res Form Appl Nat Sci.* 2013;3(1):82–7.
95. Olowu RA, Adewuyi GO, Onipede OJ, Lawal OA, Sunday OM. Concentration of heavy metals in root, stem and leaves of *Acalypha indica* and *Panicum maximum jacq* from three major dumpsites in Ibadan Metropolis, South West Nigeria. *Am J Chem.* 2015;5(1):40–8.
96. Okoye NF, Monago-Ighorodge CC, Akpobasaha NA. Evaluating the use of spiny pigweed (*Amaranthus spinosus*) and waterleaf (*Talinum triangulare*) for bioremediation of crude oil polluted soil in Ikarama Community in Bayelsa State Nigeria. *J Appl Sci Environ Manag.* 2017;21(5):903. [[CrossRef](#)].
97. Dayana PD, Manikandan K, Leninraja D, Joshep M, Kavitha Pushpam AK. Effect of saline and sodic water irrigation on physico-chemical properties of inceptisols. *Biol Forum—Int J.* 2022;14(2a):517–20.

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